



# ParaFishControl

Advancements and upcoming results

Marioletizia Fioravanti

**O.B. Dr. Ariadna Sitjà-Bobadilla**

*Project Coordinator*

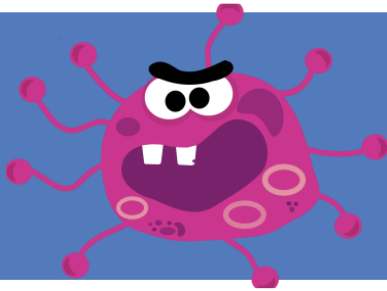


# Advanced Tools and Research Strategies for Parasite Control in European farmed fish



## 5 years collaborative project:

- Total cost: 8 104 133.75 €
- EU contribution: 7 800 000 €



## Parasites are constraining aquaculture industry

- Different parasites, culture conditions
- Different background knowledge

## Global Objective



- To increase sustainability and competitiveness of European aquaculture
- Improving understanding of fish-parasite interactions
- Developing innovative solutions and tools for the prevention, control and mitigation of the major parasites




ParaFishControl



**WHY THIS PROJECT, WHY PARASITES?**

# Economic Impact of Parasites in Finfish Aquaculture

- Direct mortality
- Morbidity: Decreased FCR & growth, parasitic castration
- Increased susceptibility to other diseases (opportunistic)
- Reduced ability to cope with changes and handling
- Harvest downgrades, loss of market, reduced durability
- Costs of treatments, prevention strategies, mort disposal, etc.
- World (Shinn et al., Global Aquaculture Advocate, 2015):
  - Estimated losses due to parasitism: hatchery (20 %), grow out (1-10% harvest)
  - Parasites' annual cost : from \$1.05 billion to \$9.58 billion
- EU (+ Norway) (H. Rodgers, FishVetGroup):
  - The value of salmon aquaculture in 2016: €12.5 billion 
  - Parasite impact: €525 to 725 million pa (direct & indirect)

# Industry



# Research



ALMA MATER STUDIORUM  
UNIVERSITÀ DI BOLOGNA



UNIVERSITÀ  
DEGLI STUDI  
DI UDINE

13  
countries

28  
partners



# WP leaders



WP1		<b>Geert Wiegertjes</b>	<a href="mailto:Geert.Wiegertjes@wur.nl">Geert.Wiegertjes@wur.nl</a>	<b>WU</b>
WP2		<b>Ivona Mladineo</b>	<a href="mailto:mladineo@izor.hr">mladineo@izor.hr</a>	<b>IOR</b>
WP3		<b>James Bron</b>	<a href="mailto:j.e.bron@stir.ac.uk">j.e.bron@stir.ac.uk</a>	<b>UoS</b>
WP4		<b>Oswaldo Palenzuela</b>	<a href="mailto:oswaldo.palenzuela@csic.es">oswaldo.palenzuela@csic.es</a>	<b>CSIC</b>
WP5		<b>Niels Lorenzen</b>	<a href="mailto:nilo@vet.dtu.dk">nilo@vet.dtu.dk</a>	<b>DTU</b>
WP6		<b>Alastair Cook</b>	<a href="mailto:alastair.cook@cefas.co.uk">alastair.cook@cefas.co.uk</a>	<b>Cefas</b>
WP7		<b>Miguel Angel Pardo</b>	<a href="mailto:mpardo@azti.es">mpardo@azti.es</a>	<b>AZTI</b>
WP8	 	<b>Marieke Reuver Emma Bello (o.b.)</b>	<a href="mailto:marieke.reuver@aquatt.ie">marieke.reuver@aquatt.ie</a>	<b>AquaTT</b>
WP9		<b>Ariadna Sitjà-Bobadilla</b>	<a href="mailto:coordination.parafishcontrol@csic.es">coordination.parafishcontrol@csic.es</a>	<b>CSIC</b>

Parasite group	Parasite species	Fish
Crustaceans	<i>Lepeophtheirus salmonis</i>	AS
	<i>Ceratothoa oestroides</i>	ESB, GSB
Monogeneans	<i>Sparicotyle chrysophrii</i>	GSB
Myxozoans	<i>Tetracapsuloides bryosalmonae</i>	RBT
	<i>Enteromyxum leei</i>	GSB
	<i>Enteromyxum scophthalmi</i>	TB
	<i>Sphaerospora molnari</i>	CC
	<i>Thelohanellus kitauei*</i>	CC
Microsporidians	<i>Enterospora nucleophila*</i>	GSB
Ciliates	<i>Ichthyophthirius multifiliis</i>	RBT, CC
	<i>Philasterides dicentrarchi</i>	TB
Dinoflagellates	<i>Amyloodinium ocellatum</i>	ESB
Amoebae	<i>Paramoeba perurans</i>	AS
Oomycetes	<i>Saprolegnia parasitica</i>	AS, RBT
Zoonotic helminths	<b>Anisakidae, Opisthorchidae, Diphyllbothriidae</b>	All

Abbreviations: AS = Atlantic salmon (*Salmo salar*); CC = common carp (*Cyprinus carpio*), ESB = European sea bass (*Dicentrarchus labrax*), GSB = gilthead sea bream (*Sparus aurata*). RBT= rainbow trout (*Onchorhynchus mykiss*), TB = turbot (*Psetta maxima*). \* Emerging or exotic parasites.



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*Enterospora nucleophila*

*Enteromyxum leei*

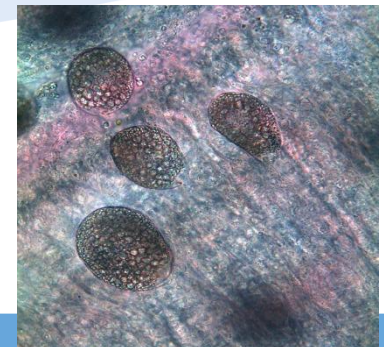


Photo by A. Siliá-Bobadilla



*Sparicotyle chrysophrii*

*Anisakis sp.*



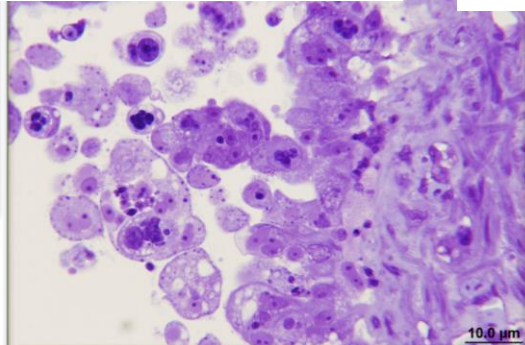
*Ceratothoa oestroides*

*Amyloodinium ocellatum*

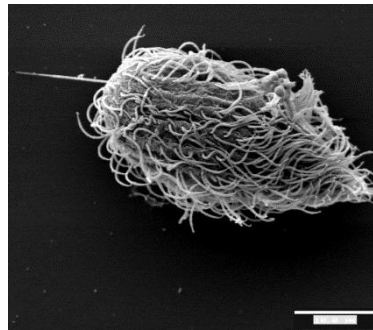




*Enteromyxum scophthalmi*



*Philasterides dicentrarchi*



*Lepeophtheirus salmonis*



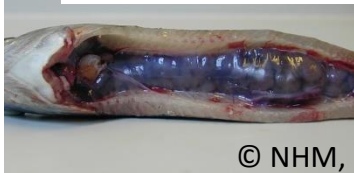
*Paramoeba perurans*



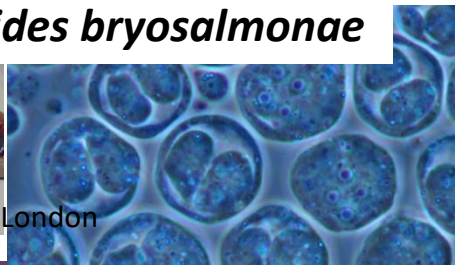
*Saprolegnia parasitica*



*Tetracapsuloides bryosalmonae*



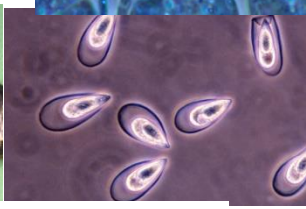
© NHM, London



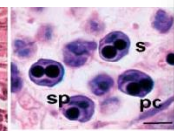
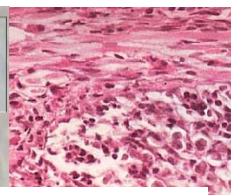
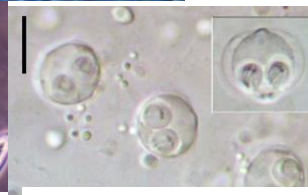
*Ichthyophthirius multifiliis*



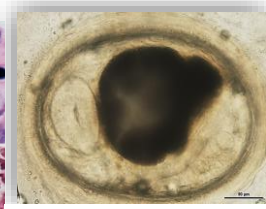
*Thelohanellus kitauei*



*Sphaerospora molnari*



© Bruno et al., 20



*Opisthorchid metacercariae*



***We are a mature project: ends 31 March 2020***

# 47 KOs collected in P3



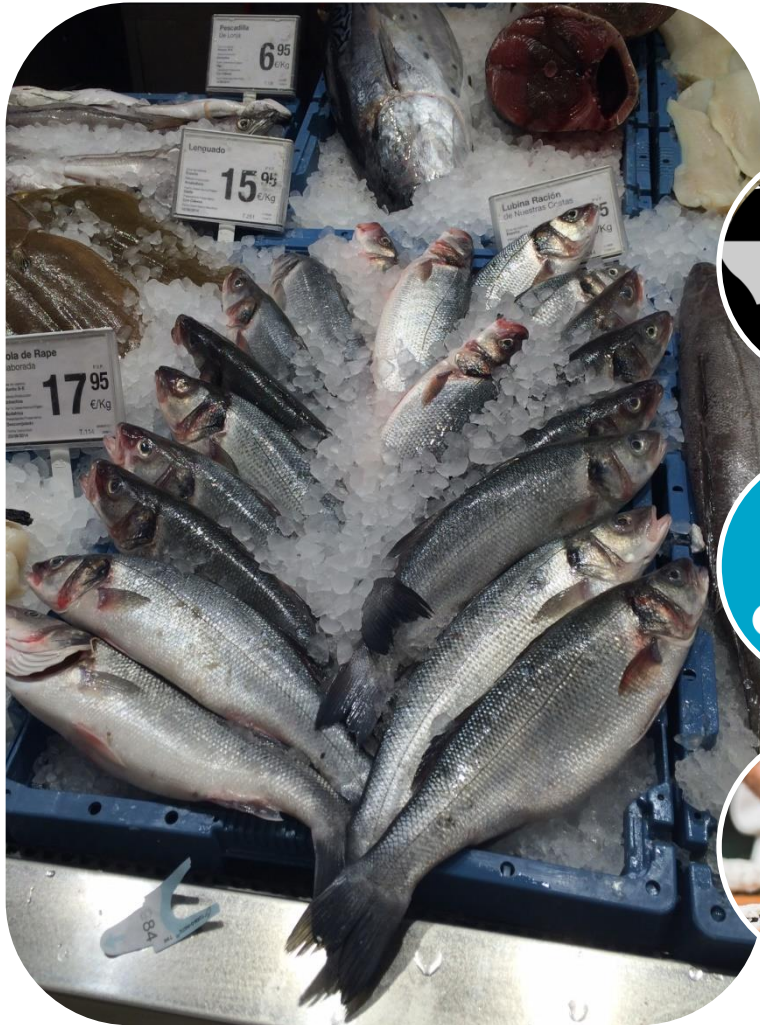
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- KO #1 - Risk Factors for parasite infections (Owner: Cefas, ANDROMEDA, AQUARK, CSIC, IOR, SKRET, USC, UNIBO, UNIUD)
- KO #2 - Infection model for *Saprolegnia* (both *diclina* and *parasitica*) (WU)
- KO #3 - White spot disease control with bacterial surfactant (Owner: KNAW, KU - Patent number EP17202669)
- KO #4 - Sparicotylosis alternative treatments (Owner: IOR)
- KO #5 - Anisakis detection portable kit (Owner: AZTI)
- KO #6 - Best Practices of farm management to avoid presence of Anisakis in farmed fish (Owner: AZTI et al.)
- KO #7 - Alternative treatment strategies for *Saprolegnia* (Owner: UNIBO)
- KO #8 - Ich recombinant protein and homogenate vaccines (Owner: KU, MTA)
- KO #9 - Expression of two proteins of *Ich* in *Pichia* (Owner: KU, W42)
- KO #10 - PKD Vaccine (Owner: UNAB)
- KO #11 - In vivo infection model for AGD in salmon (Owner: Cefas)
- KO #12 - In vitro infection model for AGD in salmon (Owner: UoS)
- KO #13 - Compound that works against *Paramoeba perurans* (AGD) (Owner: UoS)
- KO #14 - Salivary gland proteins from *Lepeophtheirus salmonis* (Owner: UiB and UoS)
- KO #15 - Vaccines against *Sphaerospora molnari* in carp (Owner: BCAS)
- KO #16 - Immunostimulants against *Sphaerospora molnari* in carp (Owner: BCAS, SKRET)
- KO #17 - qPCR for *Sphaerospora molnari* and *T. kitauei* (Owner: BCAS)
- KO #18 - Immunostimulants against *Amyloodinium ocellatum* (Owner: UNIUD)
- KO #19 - Alternative treatments for *Ceratomyxa oestroides* (Owner: IOR)
- KO #20 - Point of care for *E. leei* sea bream (Owner: CSIC, INGENASA)
- KO #21 - Infection models for *E. leei* (Owner: CSIC)
- KO #22 - Immunostimulants for *Ich* (Owner: KU)
- KO #23 - Alternative treatment strategies for Ich (Owner: MTA)
- KO #24 - Alternative treatment strategies for Ich (Owner: KU)
- KO #25 - Vaccines for *Saprolegnia* (Owner: UNAB)
- KO #26 - New monoclonal antibodies for PKD (Owner: VAL)
- KO #27 - Point of care tests for *Enteromyxum* spp. (Owner: CSIC, INGENASA)
- KO #28 - Alternative treatments for *Philasterides dicentrarchi* (Owner: USC)
- KO #29 - Vaccines for *Philasterides dicentrarchi* (Owner: USC, CSIC, INIA)
- KO #30 - Point of care test for *P. perurans* (Owner: Cefas)
- KO #31 - Repository of diagnostic methods (Owner: CSIC, DTU, UNIBO, IOR, UoS, UNAB, Cefas, BCAS, MTA, USC, KU)
- KO #32 - SHIELD new diet for sea bream (Owner: SKRET)
- KO #33 - Strategy to block the interaction of BAFF with its receptor as a treatment against proliferative kidney disease (PKD) (Owner: INIA-UNAB - Patent in progress)
- KO #34 - Challenge model for *Amyloodinium ocellatum* (Owner: UNIUD)
- KO #35 - Sequencing of *Amyloodinium ocellatum* (genome) (Owner: UNIUD)
- KO #36 - Vaccine for *Amyloodinium ocellatum* (Owner: UNIUD)
- KO #37 - Two different qPCR protocols for detection of *Neoparamoeba perurans* (AGD) (Owner: DTU)
- KO #38 - qPCR and ICH protocols for detection of *Tetracapsuloides bryosalmonae* (PKD) (Owner: DTU)
- KO #39 - qPCR for *Enterospira nucleophila* (Owner: CSIC)
- KO #40 - Methods for detection, quantification and discrimination of genotypes/serotypes of *P. dicentrarchi* (Owner: USC).
- KO #41 - Tools for detection and identification of zoonotic metacercariae (Owner: UNIBO)
- KO #42 - Sequencing of *Enteromyxum leei* (genome) (Owner: CSIC)
- KO #43 - Sequencing of *Enteromyxum scopthalmi* (genome) (Owner: CSIC)
- KO #44 - Sequencing of *P. dicentrarchi* (genome/ transcriptome) (Owner: USC).
- KO #45 - Sequencing of *Sphaerospora molnari* (genome/transcriptome) (Owner: BCAS).
- KO #46 - Sequencing of *Paramoeba perurans* (genome/transcriptome) (Owner: Cefas, UoS).
- KO #47 - Sequencing of *Sparicotyle chrysophrii* (genome/transcriptome) (Owner: IOR, HCMR, CSIC).



# 1-Generating a more positive public perception

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Environmental impact



Animal welfare



Food security

**KO#6**

**0% zoonotic helminths  
in farmed fish**



**ParaFishControl**

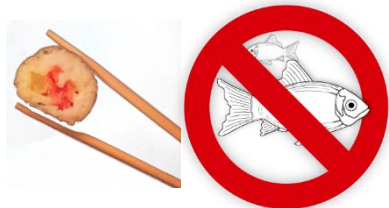


**Zoonotic risks in:**

- marine and freshwater farmed fish: 0 %
- fish products: 0%
- Feedback to EATiP, FEAP and AAC done
- Feedback to ESFA to be done



- Allergenicity of zoonotic helminths: expanded to *Contracaecum*
- Recommendation to determine the presence and transfer of allergens from aquafeeds to fish



- Smart solution to ensure safety of fish products
- Good practice handbook for Minimum Parasite Infection



We have recently presented a Spanish Standardization Proposal to the committee CTN 173/SC2 based on WP7 results

...and launched a dialogue with European authorities for an amendment to current legislation

# Interaction wild-farmed fish

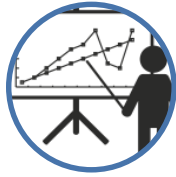
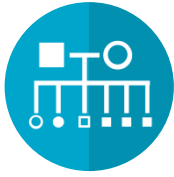


1. ***S. chrysophrii*** and ***C. oestroides*** between wild and farmed GSB and ESB
2. Characterisation of possible transfer of ***S. parasitica*** from farmed to wild salmonids and viceversa.
3. Epidemiological model to determine the transfer of ***I. multifiliis*** between farmed and wild salmonids.

The transfer depends on parasite species and microlocation (we still do not precisely which parameters, e.g. currents, bottom type, salinity...). Thus, there is a strong evidence that it does exist, but at different "success rate". It is most prominent in *Ceratomyxa* (aggressive natatory infective stages), at a lower degree for *Sparicotyle*, while *Saprolegnia* and *Ich* were evaluated using different approach, but even so.







  
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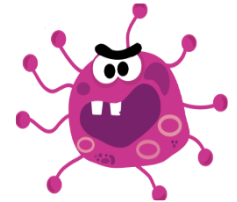


**Information essential for diagnostics, treatments, vaccines, biosecurity, breeding, welfare indicators, etc.**

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## **Knowing the parasite:**

*In vitro* culture, life cycle, experimental transmission, genome, transcriptome, proteome



## **Knowing the host:**

Immune response, histopathology, transcriptomics (RNA seq, qPCR-array)





# Some basic KOs

- KO #35** - Sequencing of *Amyloodinium ocellatum* (genome)
- KO #42** - Sequencing of *Enteromyxum leei* (genome)
- KO #43** - Sequencing of *Enteromyxum scophthalmi* (genome)
- KO #44** - Sequencing of *P. dicentrarchi* (genome/ transcriptome)
- KO #45** - Sequencing of *Sphaerospora molnari* (genome/transcriptome)
- KO #46** - Sequencing of *Paramoeba perurans* (genome/transcriptome)
- KO #47** - Sequencing of *Sparicotyle chrysophrii* (genome/transcriptome)

**KO#12** *Neoparamoeba perurans*  
RTgill-W1 In vitro challenge model

**Book** for standard Operating protocols for parasite transmission and isolation

**KO#49** *Philasterides dicentrarchi*  
*nkl* gene good candidate for genetic breeding



# KOs Vaccines and immunoprophylaxis



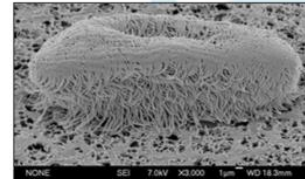
**KO #36 - *Amyloodinium ocellatum***  
Vaccine trials in lab

**KO#10 Proliferative Kidney Disease (PKD)**  
DNA Vaccine field trials

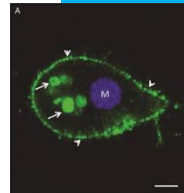
Field-based  
vaccination trials



**KO#9 *Ichthyophthirius multifiliis***  
Recombinant vaccine in *Pichia pastoris*



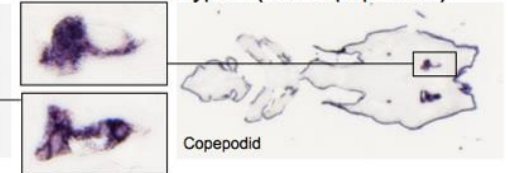
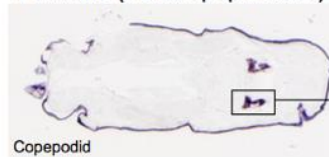
**KO#29 *Philasterides dicentrarchi***  
Variable Surface Protein (VSP)  
expressed in *Pichia pastoris* as vaccine targets



**KO#14 Salmon louse**  
Salivary gland proteins - vaccine targets

Astacins (metallopeptidases)

Trypsin (serine peptidase)



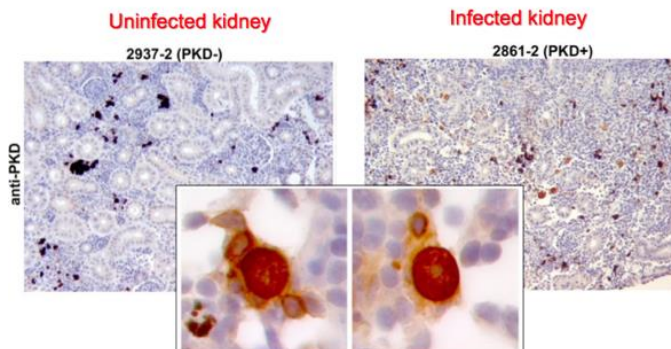
***Enteromyxum leei***  
functional feeds  
And others "in the oven"...



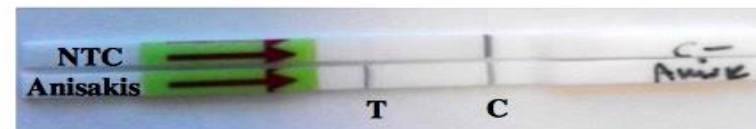
# KOs diagnostic tools



## **KO#26** Proliferative Kidney Disease (PKD) P14G8 Mab



## **KO#5** *Anisakis* Portable detection kit



LF RPA Assay for the detection of *Anisakis* simplex: Test line (T); Internal Control line (C). Negative Template control (NTC)

## **KO #40** *P. dicentrarchi* Methods for detection, quantification and discrimination of genotypes/serotypes

## **KO#41** Zoonotic metacercaria Tools for detection and identification

## **KO#39** *Enterospora nucleophila* qPCR & ISH protocols

**POC:** *Enteromyxum* spp.

**Reference diagnostic methods** for: *P. perurans*, *E. nucleophila* *T. bryosalmonae*

**Repository** of parasite diagnostic methods

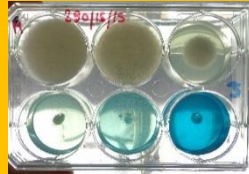


# KOs treatments



## **KO#7** *Saprolegnia* Treatments validation

Benzoic acid (MIC:100 ppm)  
Acetic acid (MIC:250 ppm)  
Iodoacetic acid (MIC:250 ppm)  
copper sulphate (MIC:250 ppm)  
Virkon™S (MIC = MLC = 1,000 ppm)  
Actidrox® (MIC = 5,000 / MLC= 500)  
Detarox® AP (MIC = 1,000 ppm / MLC= 100)



## **KO#4** Sparicotylosis Alternative Treatments

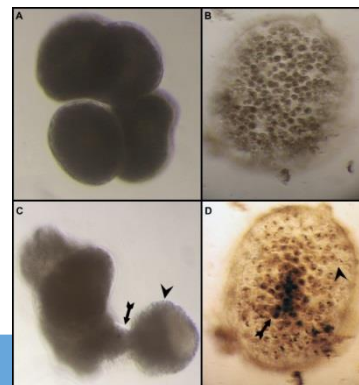
Bithionate sodium  
Cedrol  
Curcumin  
Eucalyptol  
Garlicin  
(+)-trans-chrysanthemic acid  
Camphor

## **KO#28** *Philasterides dicentrarchi* Nk-lysin (Nkl) AMP

**KO #33** PKD Strategy to block the interaction of BAFF with its receptor as a treatment (patent in progress)

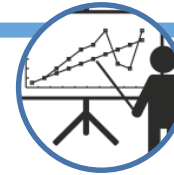
Optimal strategy for the use of salmon lice cleaner fish

## **KO#3** *Ichthyophthirius multifiliis* Bacterial Bio-surfactant (*Pseudomonas*)



PATENT  
EDP ref: P1600045PC00 –  
International PCT Application No.  
PCT/EP2018/081923

START-UP: SUNDEW



Risk  
Factor  
Analysis

**KO#7** Identification of risk factors for parasite introduction into and amplification within fish farms: *Sparicotyle chrysophrii*, *Enteromyxum leei*, *Enterospora nucleophila*, *Ceratomyxa oestroides*

## Alpha version of the economic model

- Farm level tool developed in R Shiny
- Provides a user-friendly web-based interface through which partners and fish farmers can investigate some of the key findings
- Provides information on the relative likelihood of infection and intensity based on factors chosen by an end user.



Manuals describing IPMS for parasite management and handbooks describing good practices

- **Manual 1 - Salmonids**
- **Manual 2 - European sea bass & gilthead sea bream**
- **Manual 3 - Turbot**
- **Manual 4 - Common Carp**



**Integrated  
Parasite  
Management  
Strategies  
(IPMS)**



# Next event



## FINAL CONFERENCE BRUSSELS

11<sup>th</sup> March 2020

CSIC Headquarters

Registration: [bit.ly/2NO1NrO](https://bit.ly/2NO1NrO)



Contact us



## Communication & Press

- Emma Bello
- emma@aquatt.ie



## Management

- Dr. Enric Belles-Boix
- enric.belles-boix@inra.fr



## Coordination

- Dr. Ariadna Sitjà-Bobadilla
- parafishcontrol.coordination@csic.es



- <http://www.parafishcontrol.eu>





