

Gene editing by CRISPR/Cas9: A game changer for aquaculture?

Lene Kleppe

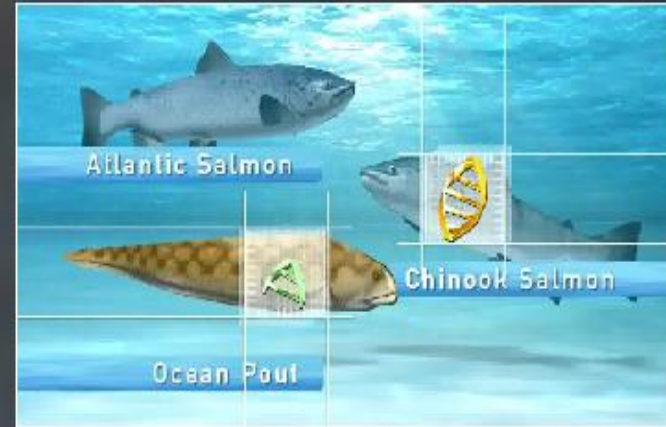
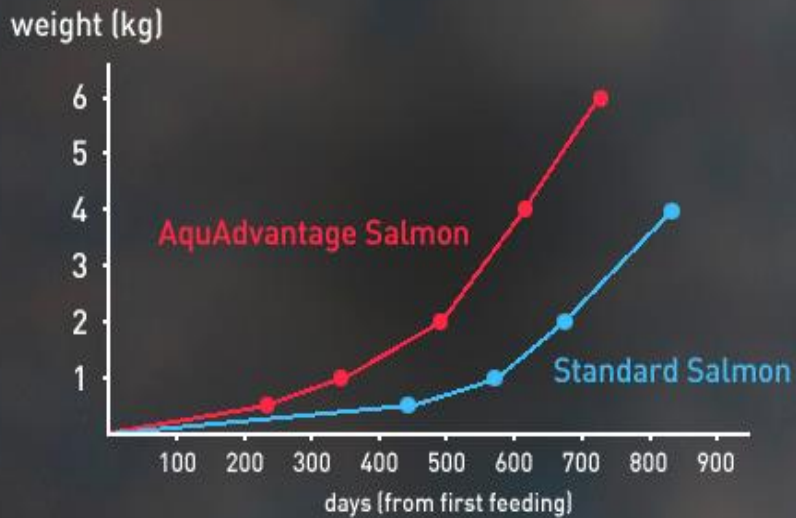
Institute of Marine Research, Norway



Institute of Marine Research, Norway



Transgenesis



AquAdvantage salmon are Atlantic salmon with a growth hormone gene from chinook salmon, to accelerate growth, and a fragment of DNA from ocean pout, to help activate the chinook gene.

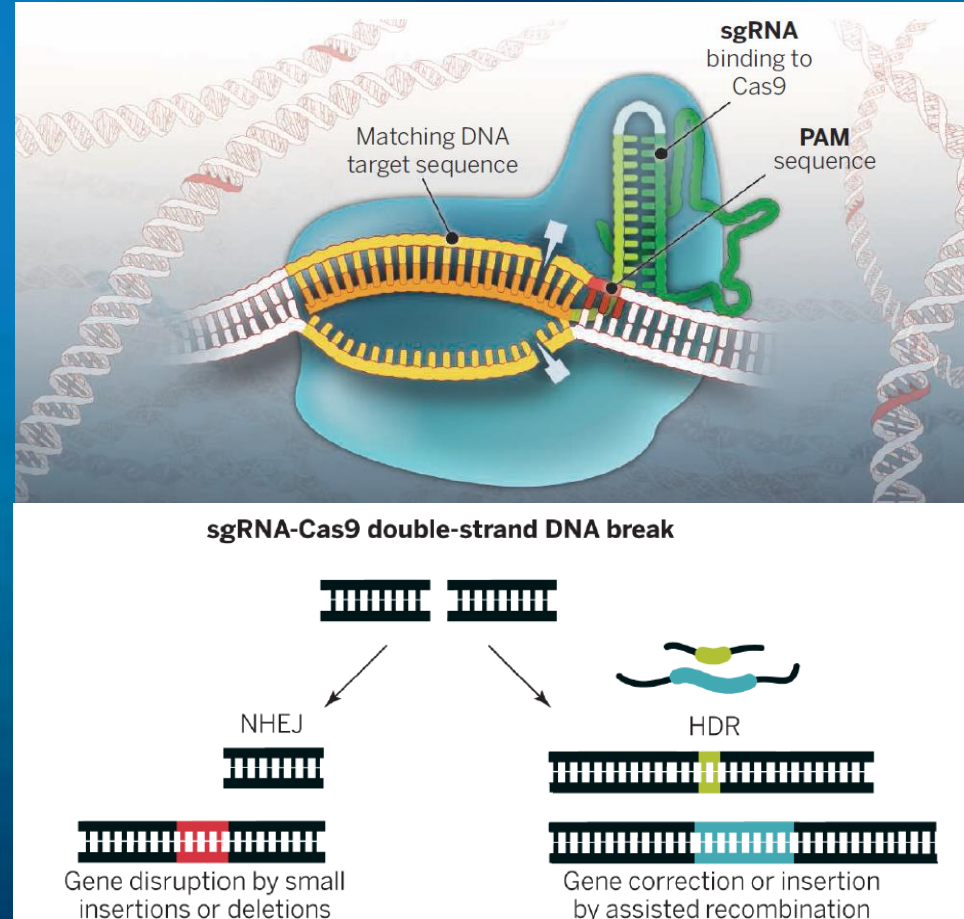


Source: Images, video and chart data courtesy of AquaBounty Technologies



Gene editing by CRISPR/Cas9

Clustered Regularly Interspaced Short Palindromic Repeats/CRISPR associated protein 9



NHEJ=
Non-Homologous End Joining

HDR=
Homology Directed Recombination



CRISPR/Cas9

Research tool and possible industrial applications

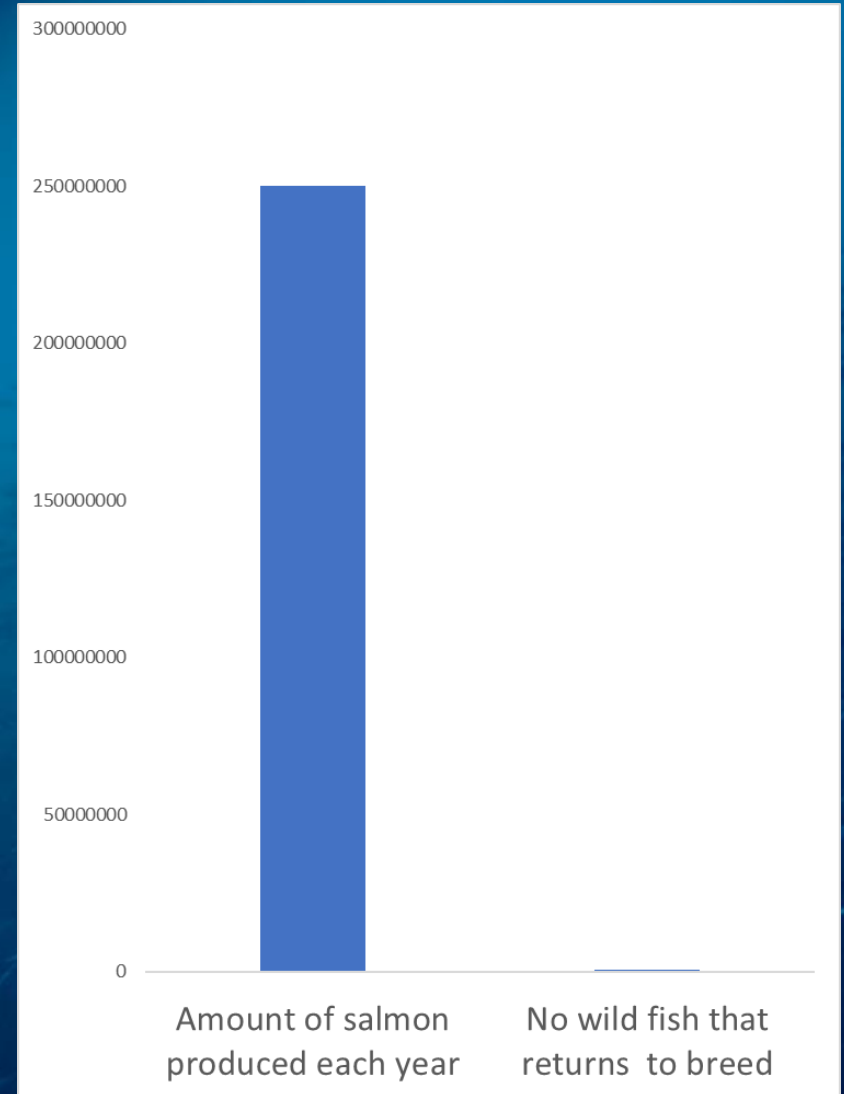
- Genetic containment of wild populations (salmon)
- Filet quality (salmon)
- Disease resistance (pig)
- Welfare (cow)
- Move traits between strains (salmon)



Environmental challenges – escaped salmon

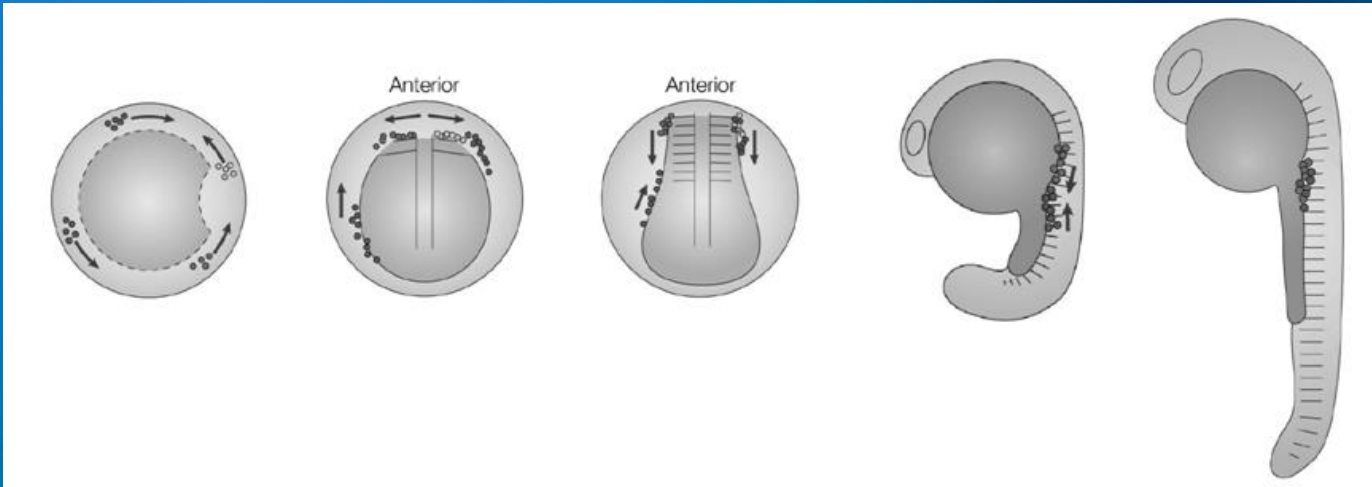
Genetic introgression with wild populations

Solution: sterile salmon

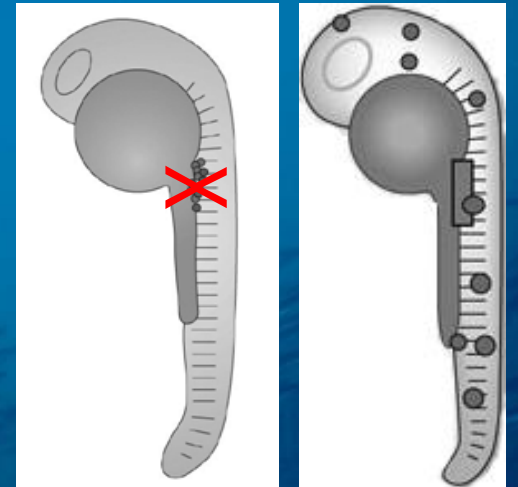


Genetic containment of wild populations

Sterile salmon through germ cell ablation



Disrupt the migration of primordial germ cells



No germ cells will form
= sterility

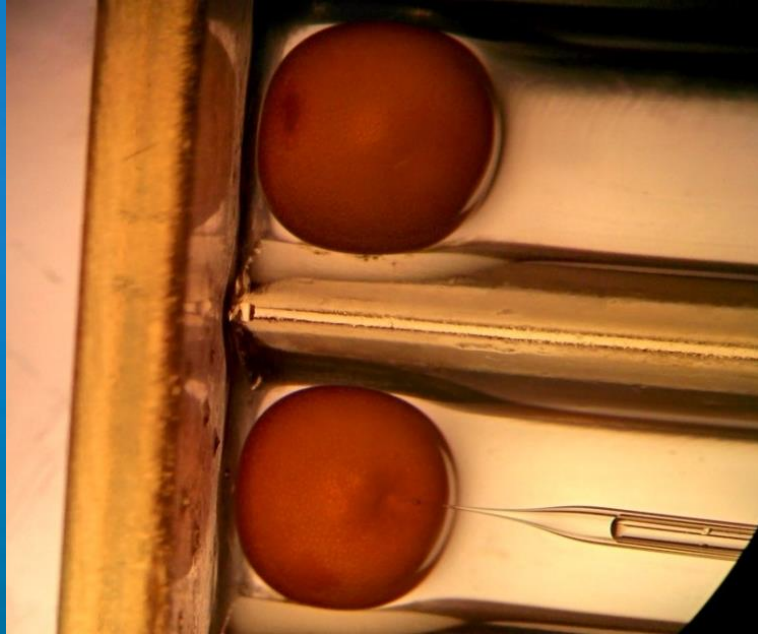
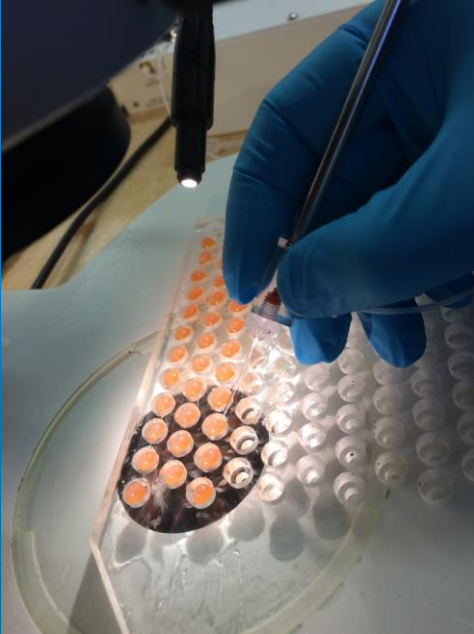
Dead end (*dnd*) gene:

- RNA binding protein
- *dnd* mutation in mice
- *dnd* knockdown in fish
(zebrafish, medaka, pond loach, rainbow trout, starlet)

Lack of primordial germ cells



Exploring the CRISPR/Cas9 methodology in salmon



Injection of guide RNA + Cas9 in newly fertilized eggs
Targeting a pigmentation gene: *slc45a2* (*albino*)

Successful editing = albinos

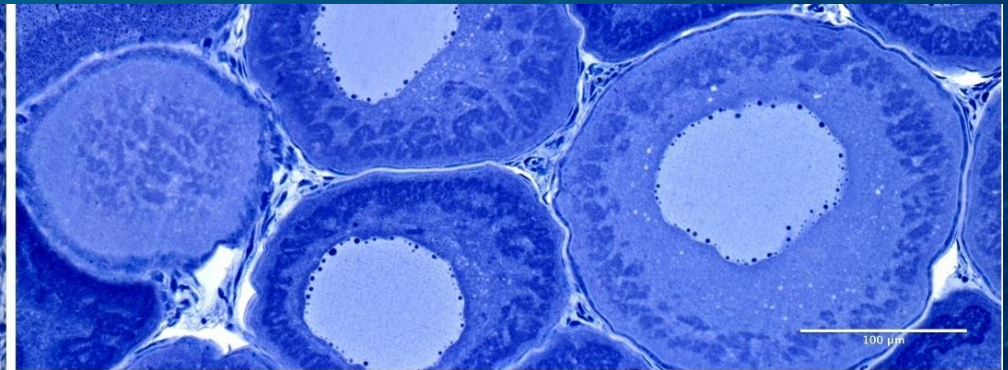
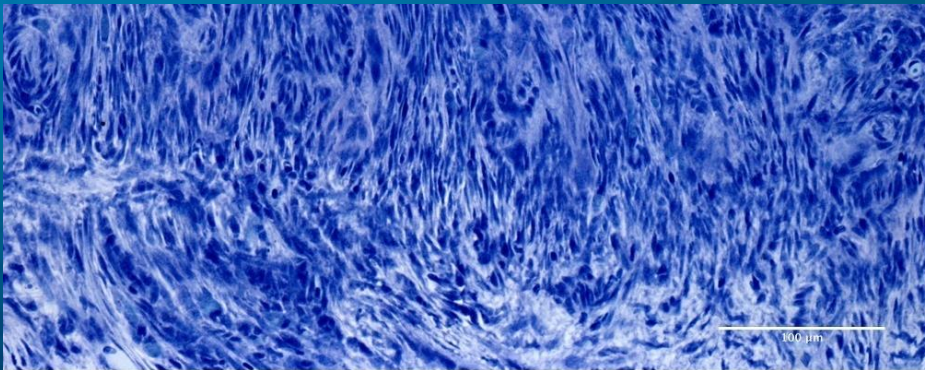
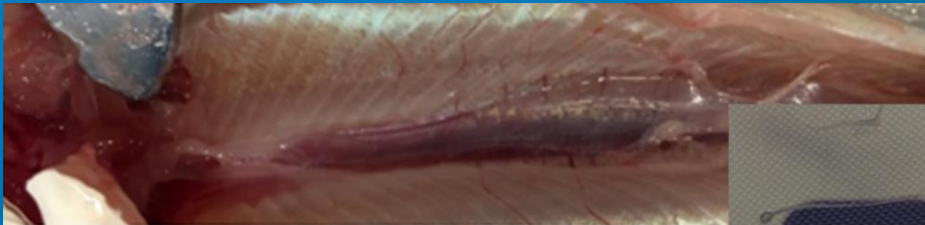


Germ cell-free (sterile) salmon

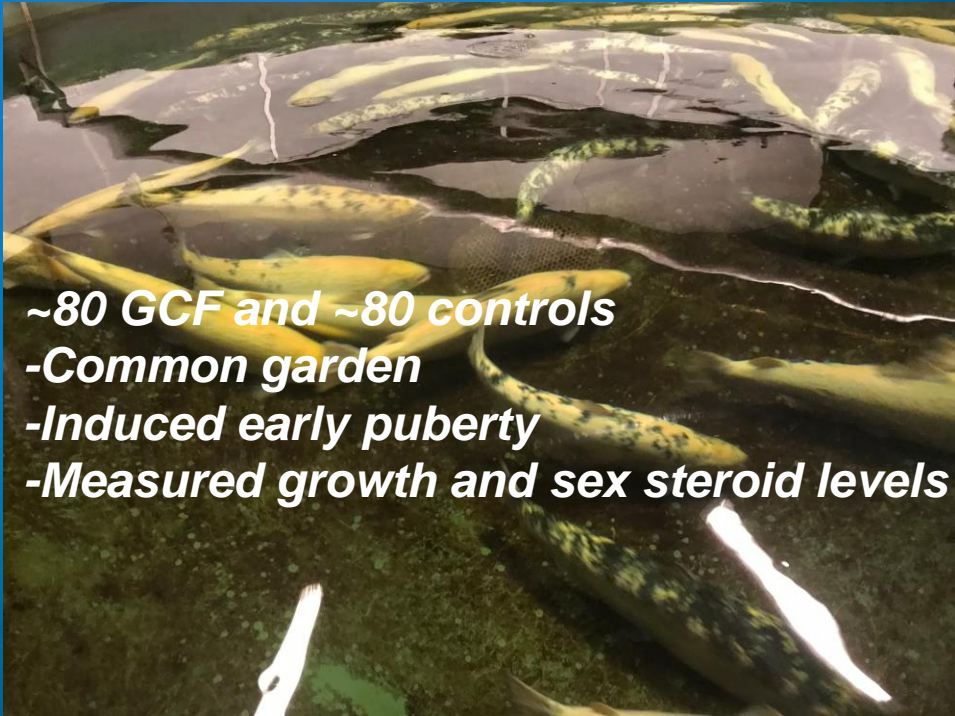
dnd + albino Knockout female



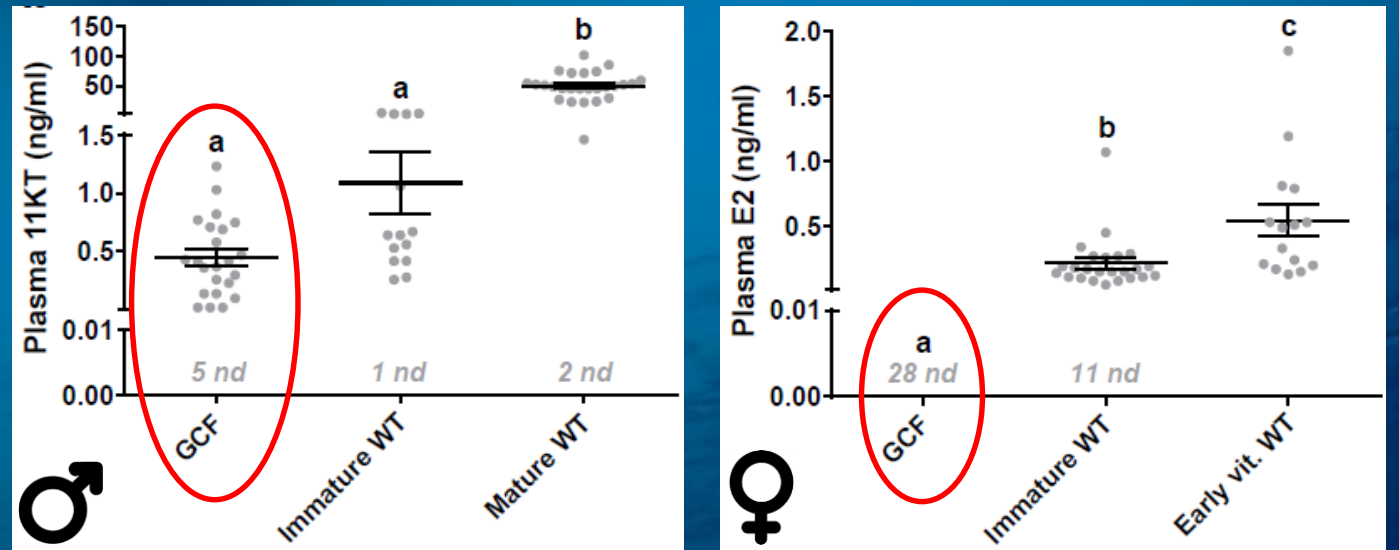
Control female



Germ cell-free (GCF) salmon



Growth and sexual maturation



Very low or no production of sex steroids



Ongoing: New, long-term growth experiment

Environmental challenges – feed resources

Marine resources are still needed to produce salmon feed

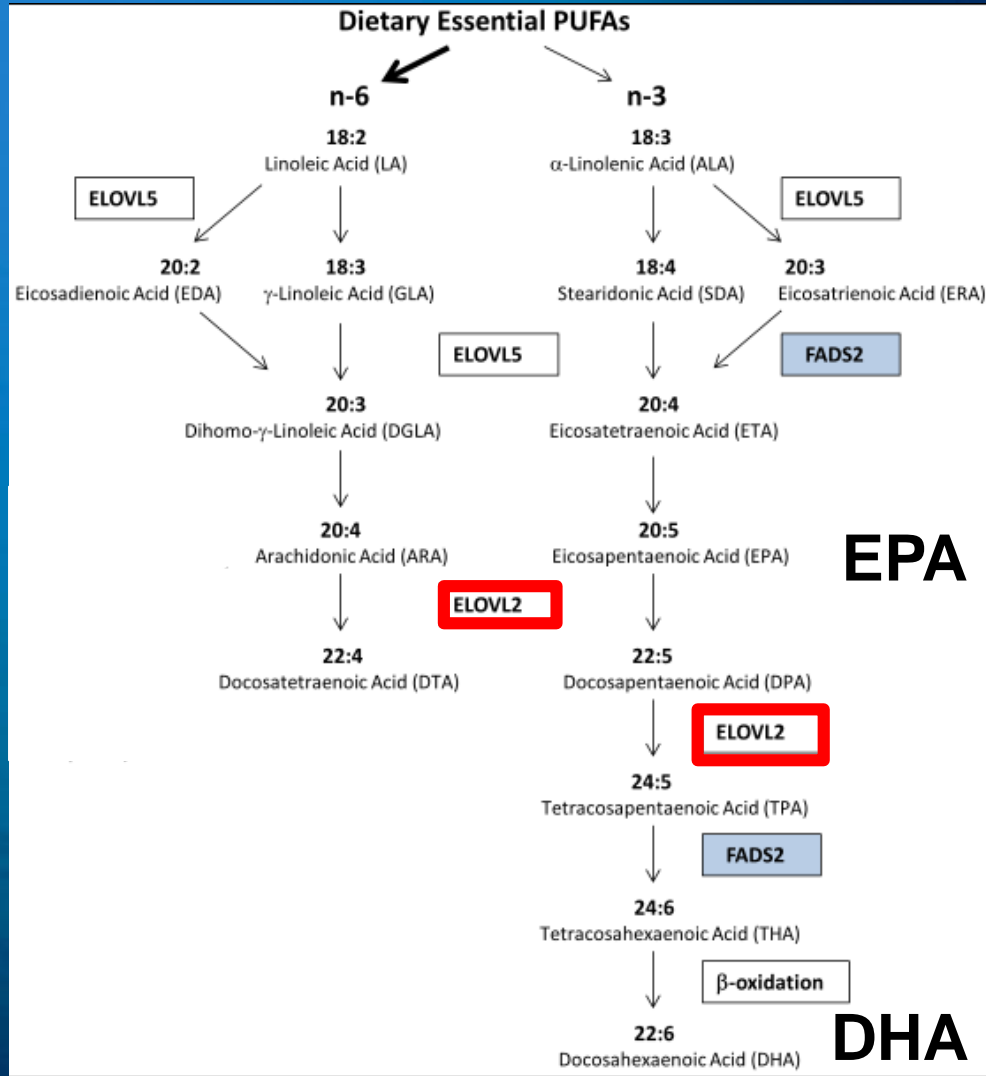
Increasing proportions of plant based ingredients
→ Less omega-3 fatty acids both in feed and filet



Can we make the salmon produce more of its own omega-3 fatty acids?



Synthesis of omega-3



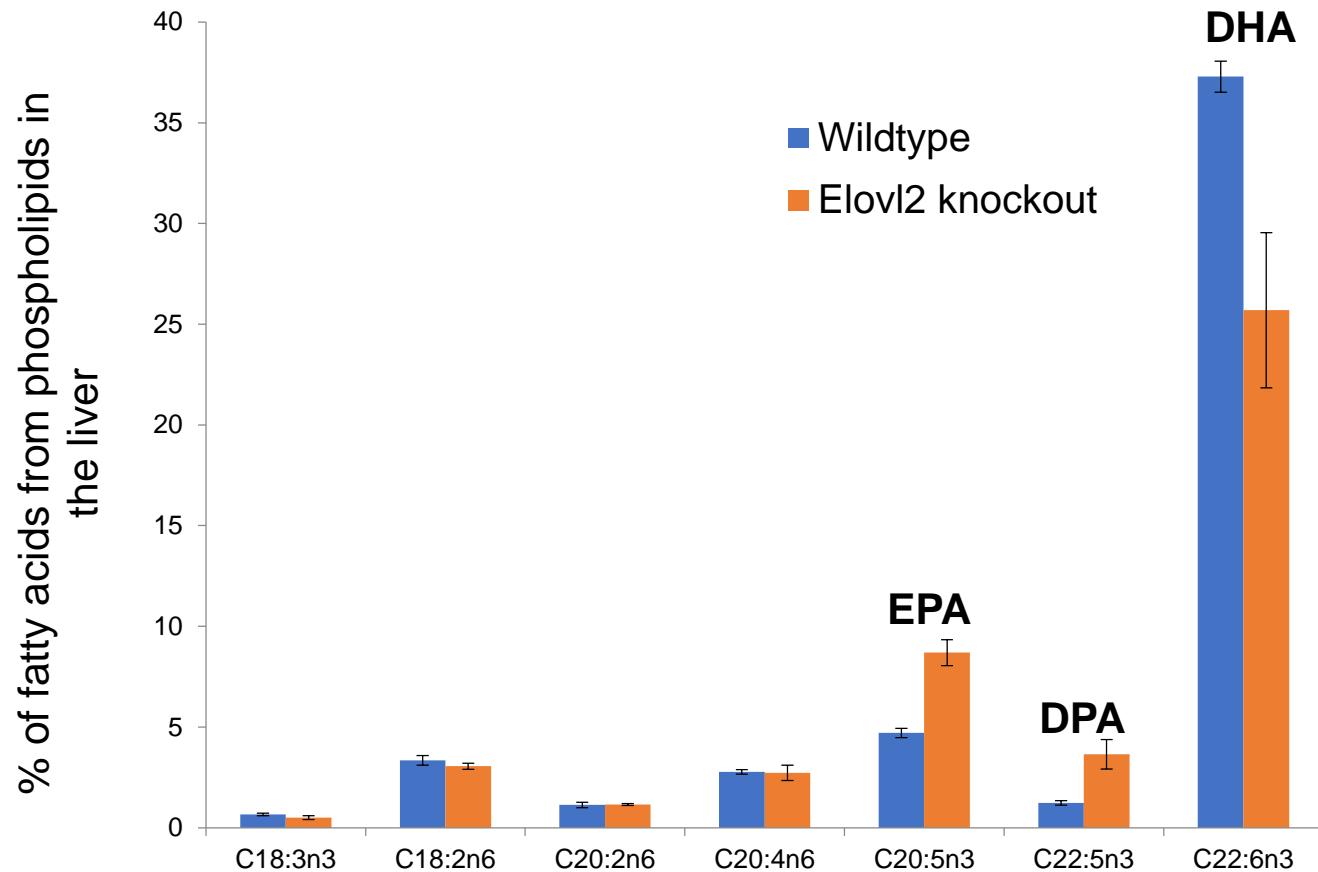
Control



elovl2 + albino knockout



Synthesis of omega-3



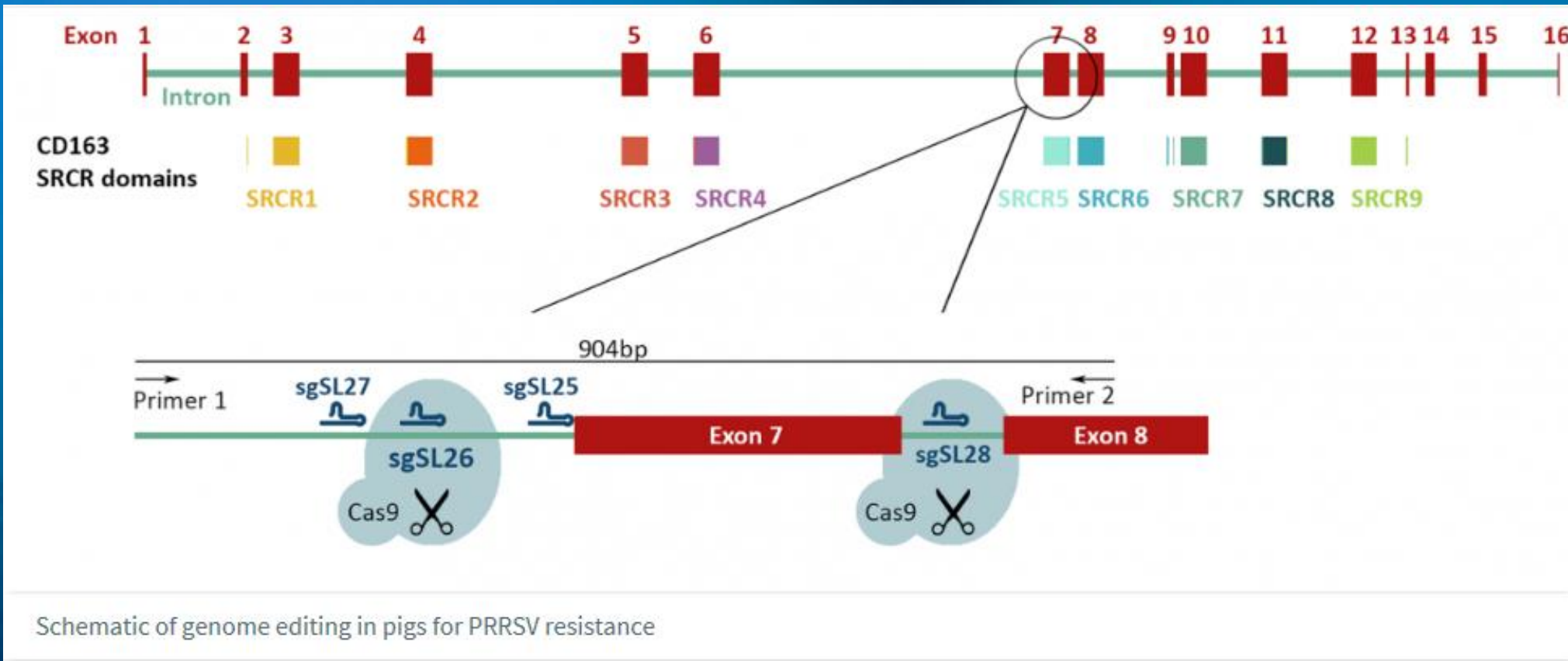
elovl2 knockout inhibits elongation of EPA and DPA

	C20:5n3	C22:5n3	C22:6n3
p-values	0,0043	0,027	0,031

Prrsv resistant pig

Porcine reproductive and respiratory syndrome virus (PRRSV)

costs the United States swine industry around \$644 million annually, and in Europe about 1.5b€ every year.



The virus infects macrophages

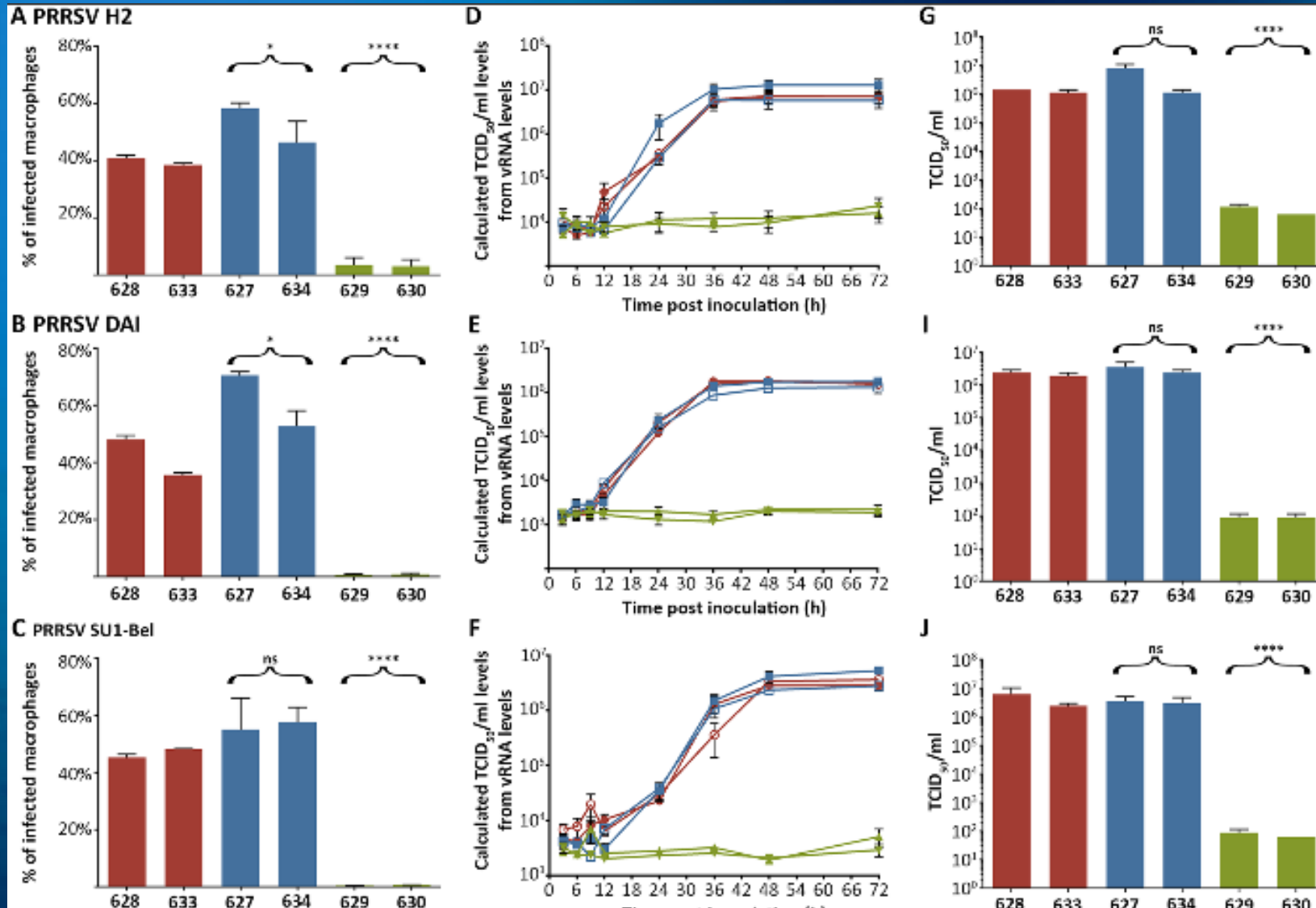
SRCR5 domain: Interaction site for the virus

CD163 has several functions

Schematic of genome editing in pigs for PRRSV resistance



Prrsv resistant pig



Control
Heterozygous
Biallelic/homozygous

H2, DAI and SU1-Bel
 = subtypes of the virus

Pulmonary alveolar macrophages
 were collected from lungs, and
 infected with PRRSV.

-% infected macrophages (A-C)
 -Virus replication growth (D-F)
 -Conc. of infectious viral particles
 (G-J)

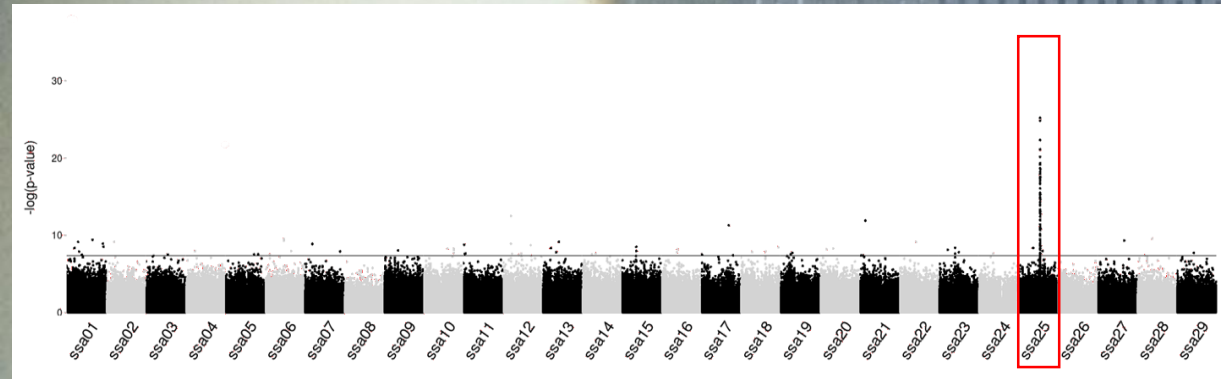
Knock in of wild traits beneficial for salmon aquaculture

- Disease resistance
- Sea water adaptation
- Nutritional content of filet
- Time of maturity

Move traits between salmon strains

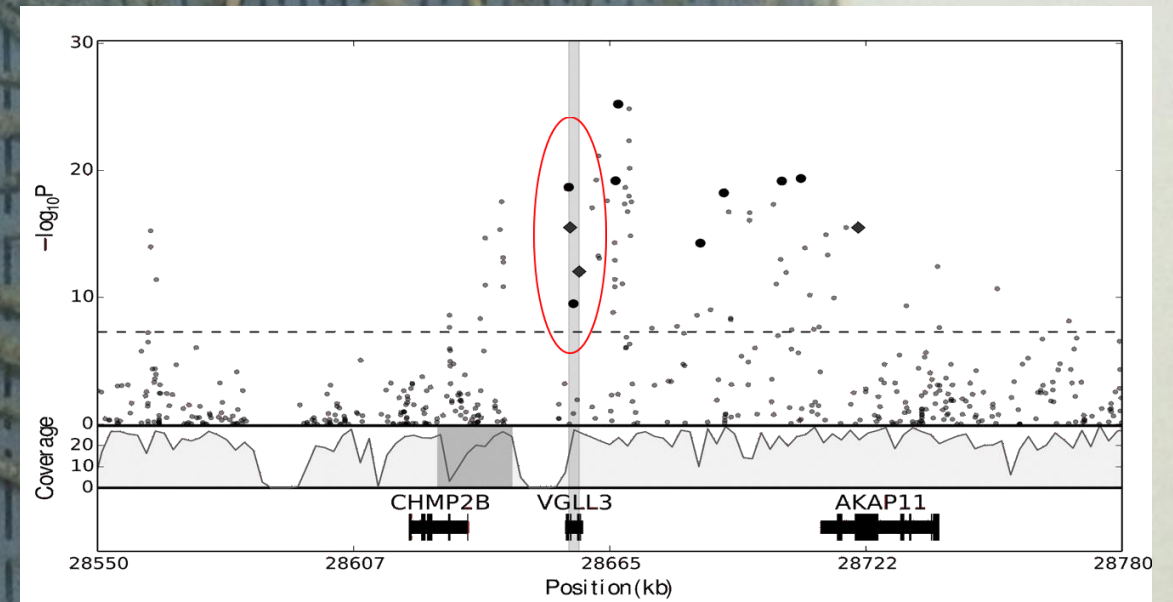
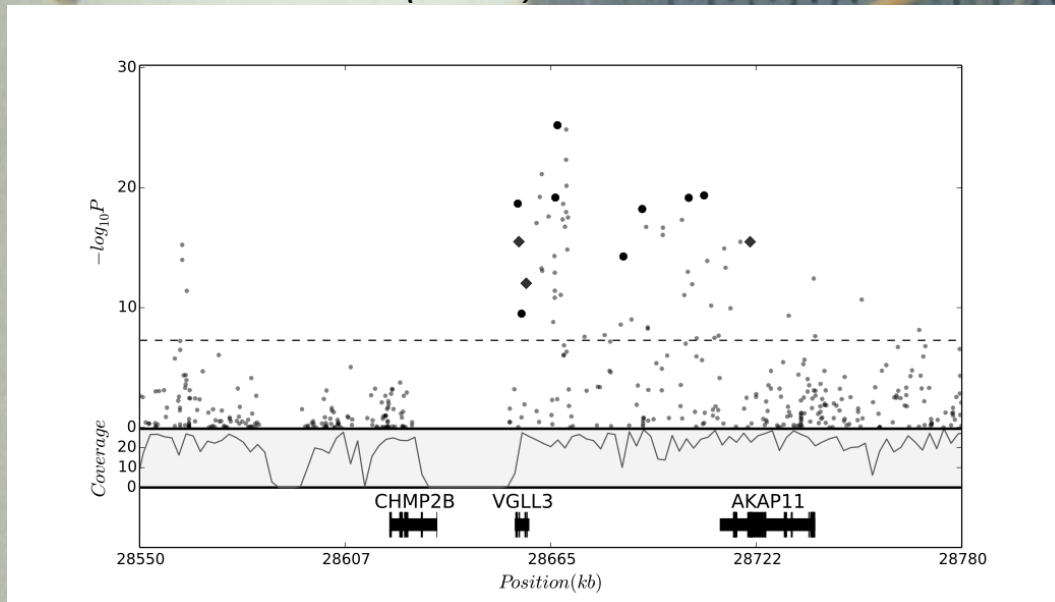
Knock in of wild traits beneficial for salmon aquaculture

- Disease resistance
- Sea water adaptation
- Nutritional content of filet
- Time of maturity



Ayllon et al., Plos Genetics (2015)

Barson et al., Nature (2015)



Move traits between salmon strains

Potential use of CRISPR in aquaculture

A scenic view of a fjord in Norway, with snow-capped mountains in the background and a fish farm (cage culture) in the foreground. The water is calm, reflecting the surrounding landscape. The fish farm consists of several long, narrow cages extending into the water, with various structures and equipment visible on the floating platforms.

Benefits

- Precise and fast targeting of genes
- Can knock out, correct or add DNA
- Less risk/ethical consideration than classical transgenic methods
(do not need to mix DNA from different species)

Risks

- Off-target activity (solved by sequencing)
- Genetic introgression of farmed fish into wild populations
(solved by genetic containment)
- Trust

This work was carried out by:



Eva Andersson, Sven Leining, Kai Ove Skaftnesmo, Geir Lasse Taranger, Anna Wargelius, Rolf Brudvik Edvardsen, Lene Kleppe



Geir Dahle



Vidar Wennevik



Monica Solberg



Kevin Glover



Simon Lillico, Jonathan Pavelin, Christopher Proudfoot, Ross Houston



Carl Johan Rubin



Fernando Ayllon



Erik Kjærner-Semb



Tomasz Furmanek



Rüdiger W. Schulz



Tom Hansen



Alex Datsomoor



Per Winge



Dorothy Dankel



Birgitta Norberg



Per Gunnar Fjeldal





Thank you for the attention!

Thanks to Norwegian Research Council and EU for funding this research.
NRC BIOTEK2021 project SALMOSTERILE , HAVBRUK_BIOTEK2021 project SALMAT
HAVBRUK project STERWELL and MATGEN
EU- AQUACRISPR